

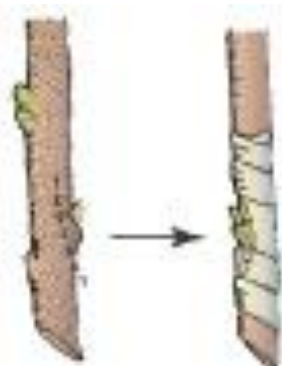
Berry viruses complexes and what we can do about it

Ioannis E. Tzanetakis

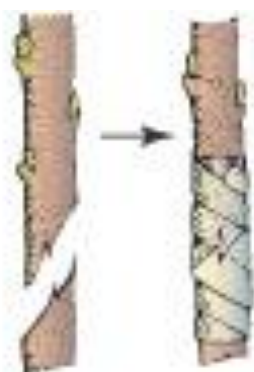
itzaneta@uark.edu
479-575-3180

Virus transmission 101

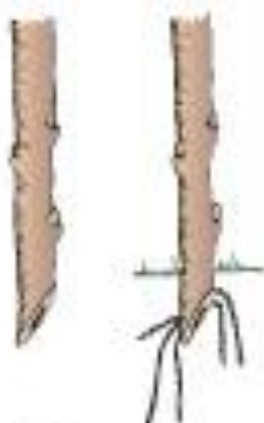




By budding



By grafting



By cuttings



By bulbs



By corms



By rhizomes



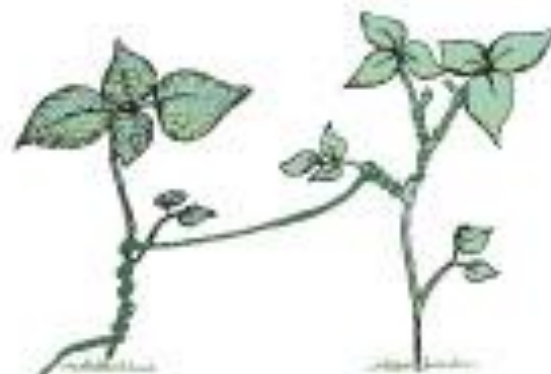
By tubers



By runners (stolons)

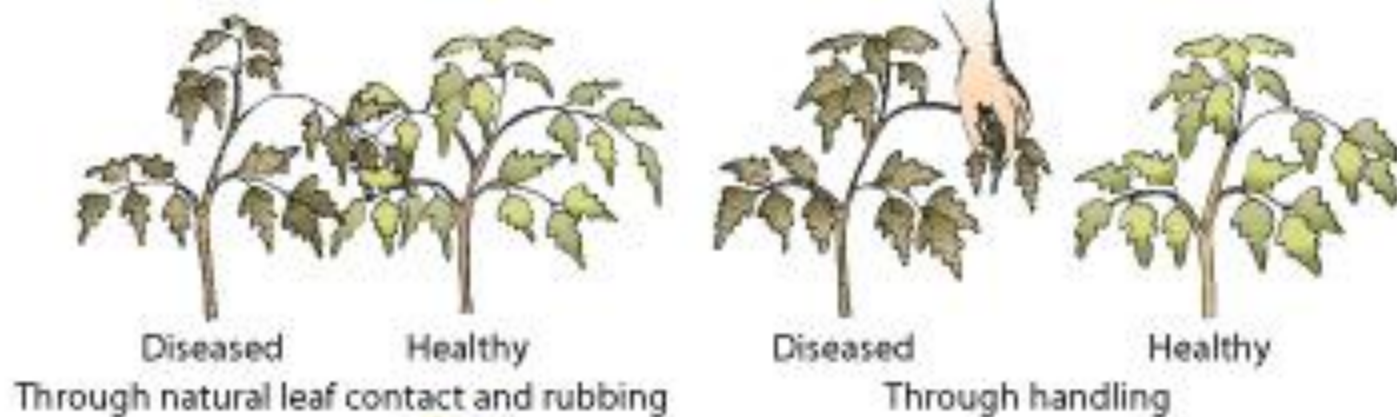


Through natural root grafts

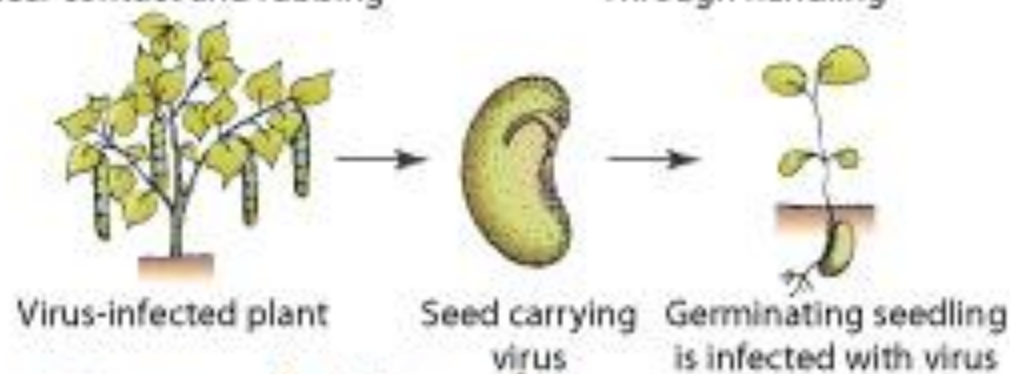


Through dodder

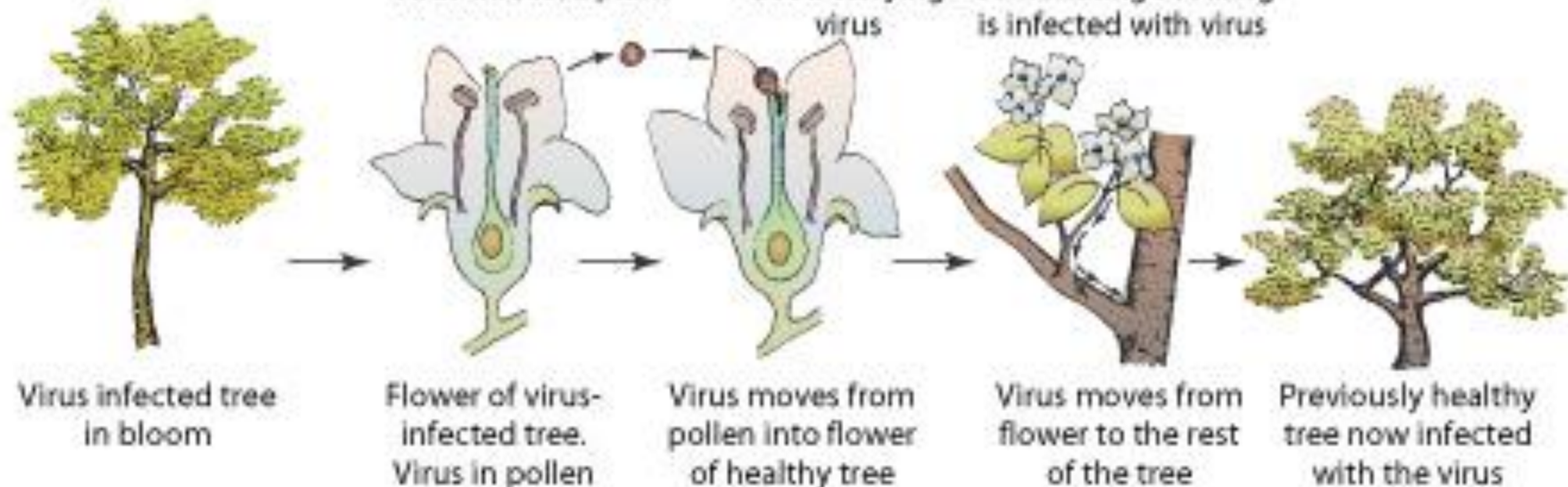
Through contact



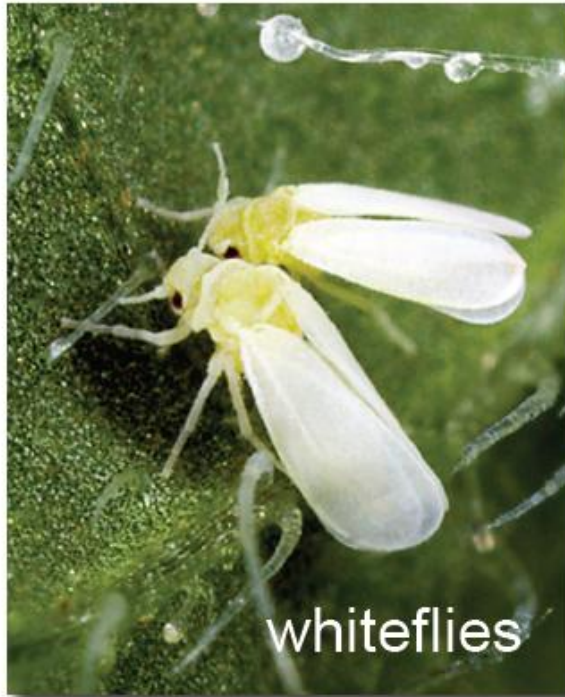
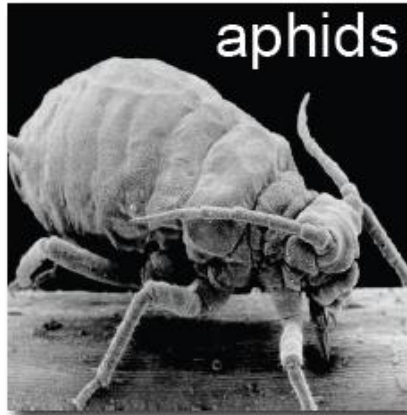
Through seed



Through pollen



Phylum *Arthropoda*, Class *Insecta*, Order *Hemiptera*



Phylum *Arthropoda*, Class *Insecta*

Order *Thysanoptera*



Order *Coleoptera*

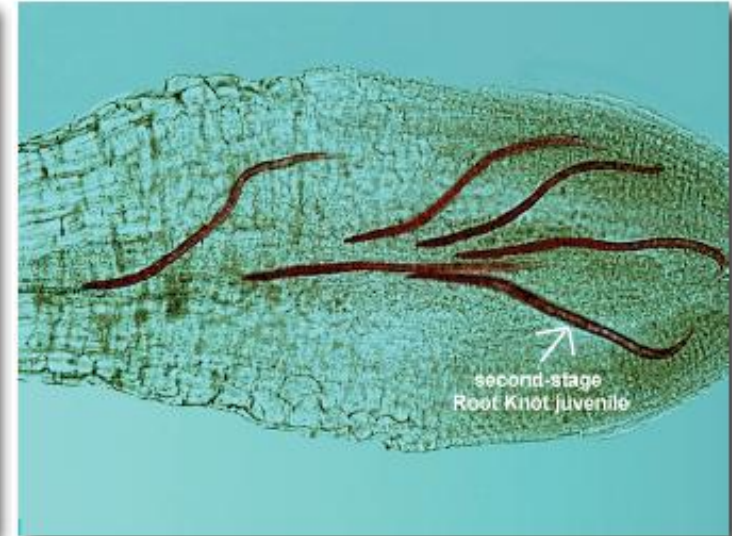


Phylum *Arthropoda*, Class *Arachnida*, Order *Acariformes*

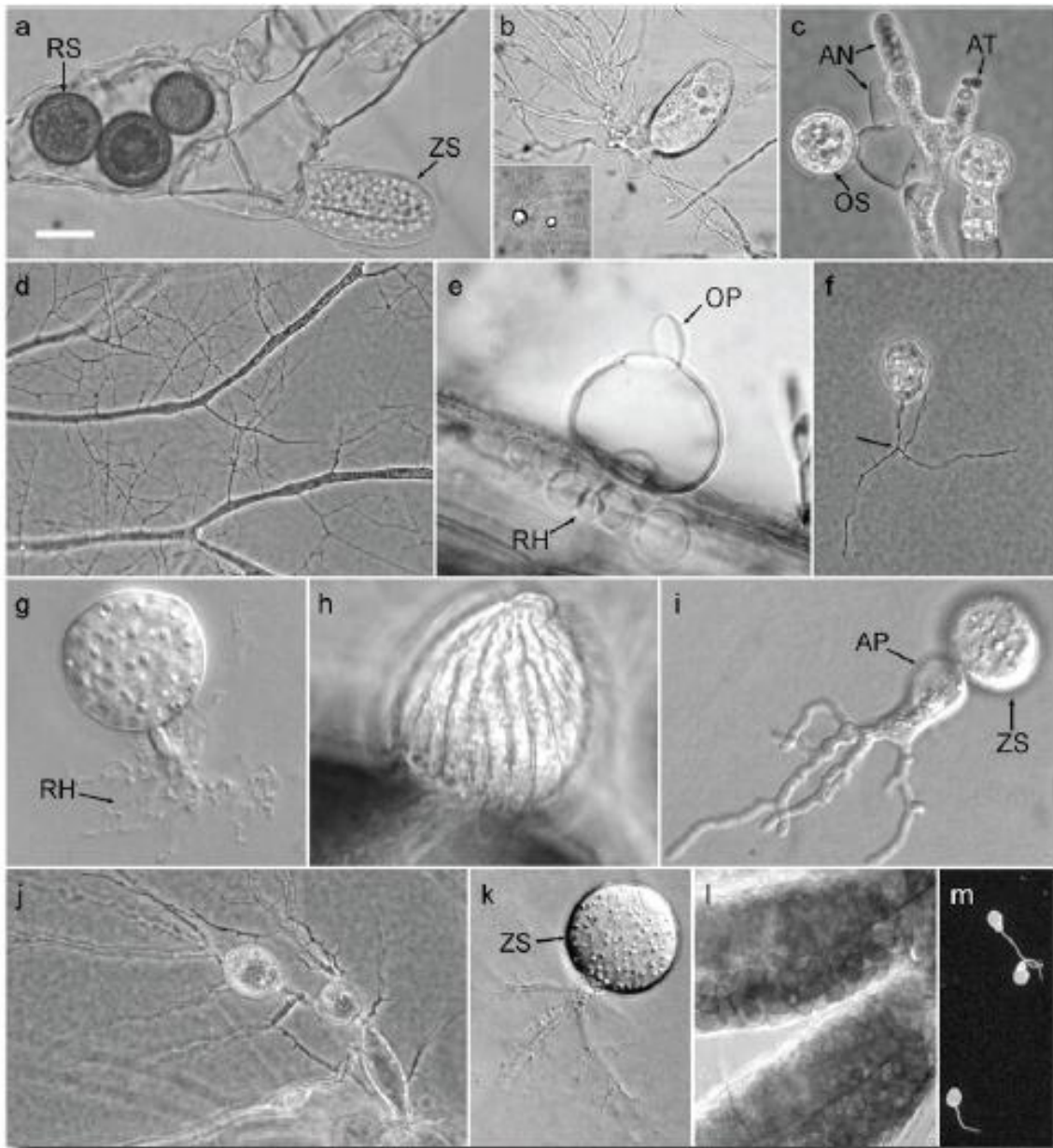


mites

Phylum *Nematoda*, Class *Secernentea*, Order *Tylenchida*



Nematodes



Kingdom: *Fungi*

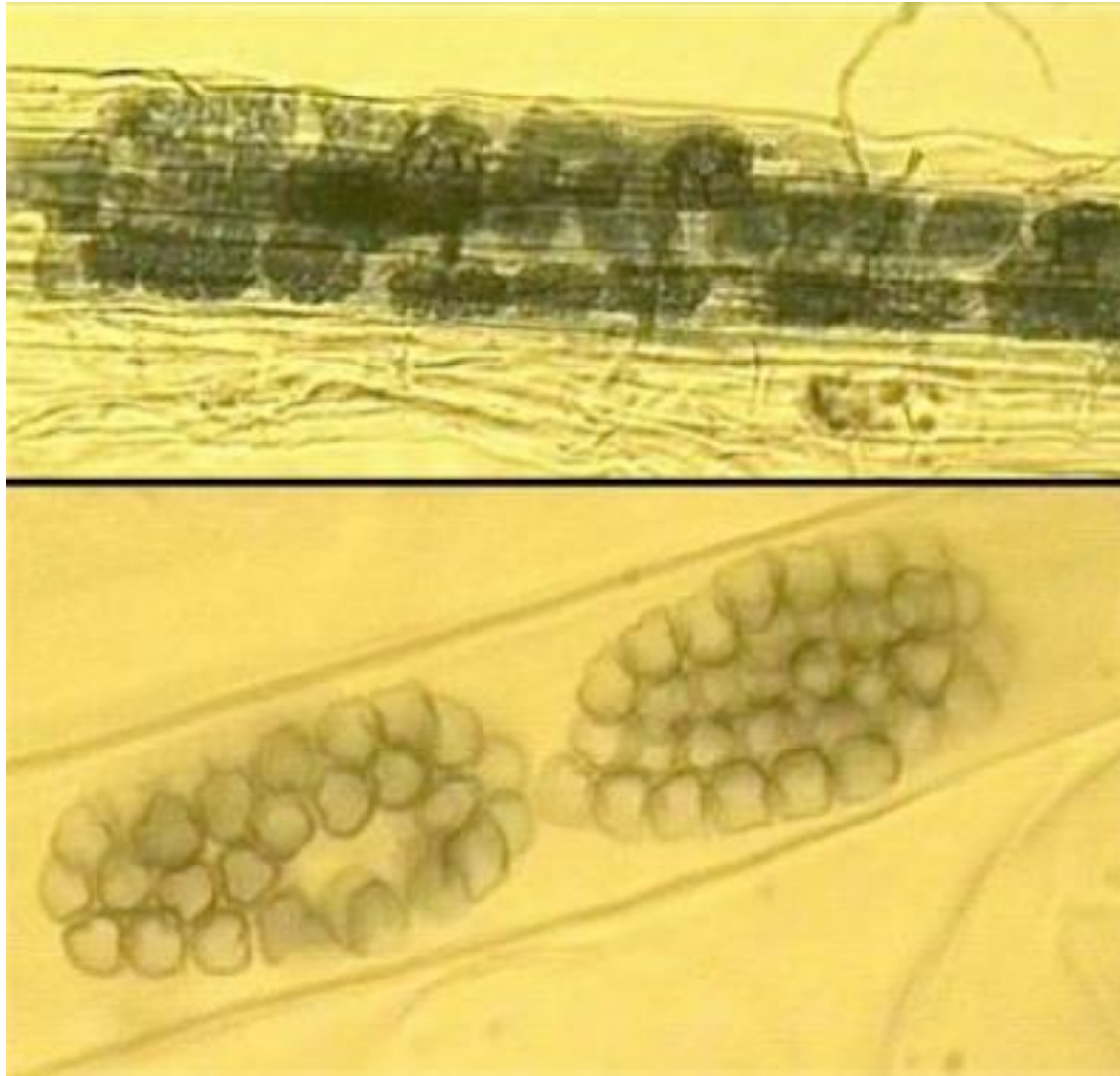
Phylum: *Chytridiomycota*

Class: *Chytridiomycetes*

Order: *Uncertae sedis*

Family: *Olpidiaceae*

Genus: ***Olpidium***



Kingdom: *Rhizaria*

Phylum: *Cercozoa*

Class: *Plasmodiophorea*

Order: *Plasmodiophorida*

Family: *Plasmodiophoridae*

Genus: ***Polymyxa***

Vectors and plant viruses they transmit

Vector taxa	Vector group	Virus groups				Total	%
		Icosahedral particles RNA genome	Rod-shaped particles RNA genome	DNA genome	Enveloped particles RNA genome		
Hemiptera	Aphids	26	153 ^a	13	5	197	28
	Whiteflies	–	13	115 ^b	–	128	18
	Leafhoppers	8	–	15	3	26	4
	Planthoppers	10	4 ^c	–	4	18	3
	Other hemiptera	–	8	5	–	13	2
Thysanoptera	Thrips	2	–	–	14	16	2
Coleoptera	Beetles	50	1	–	–	51	7
Acari	Mites	10	9	–	–	10	1
Nematoda	Nematodes	45	3	–	–	48	7
Mycota	Fungi	8	16	–	–	24	3
	No identified vectors	84	60	19	3 ^d	166	24
	Total	233	268	167	30	697	
	%	33	39	24			

^aIncludes 110 virus species of the genus *Potyvirus*, family *Potyviridae*;

^bVirus species of the genus *Begomovirus*, family *Geminiviridae*;

^cThese are all tenuiviruses that have multiple shapes;

^dThese viruses probably have insect vectors.

Four modes of virus transmission

Biological characteristic	Nonpersistent stylet-borne	Semipersistent foregut-borne ^b	Persistent circulative	Persistent propagative
AAP and IAP ^a	Seconds, minutes ^c	Minutes, hours ^d	Hours, days ^d	Hours, days ^d
Latent period	None	None	Hours, days	Days, weeks
Retention time in vector	Minutes, lost after molting	Hours, lost after molting	Days, weeks	Lifespan of insect
Presence in vector's hemolymph	No	No	Yes	Yes
Multiplication in vector	No	No	No ^e	Yes
Transovarial transmission	No	No	No	Often

^aAAP, Acquisition access period; IAP, Inoculation access period;

^bA recent publication revealed that the semi-persistent virus *Cauliflower mosaic virus* (CaMV) is retained in the stylet (178);

^cThe time period during which virus can be acquired from and inoculated into plant epidermal cells;

^dAAP and IAP times depend on the location of the virus in the plant, i.e., acquisition of the virus from the plant phloem takes longer than acquisition from the epidermis or mesophyll cells;

^eExcept for TYLCV for which there is evidence that it replicates in its whitefly vector.

Strawberry viruses circa 2003

Arabis mosaic – Europe

Tomato ringspot

Raspberry ringspot – Europe

Strawberry latent ringspot-Europe

Tomato black ring –Europe

Nematodes

Strawberry crinkle

Strawberry mild yellow edge

Strawberry mottle

Strawberry vein banding

Aphids

Fragaria chiloensis latent – Chile

Tobacco streak*

Pollen

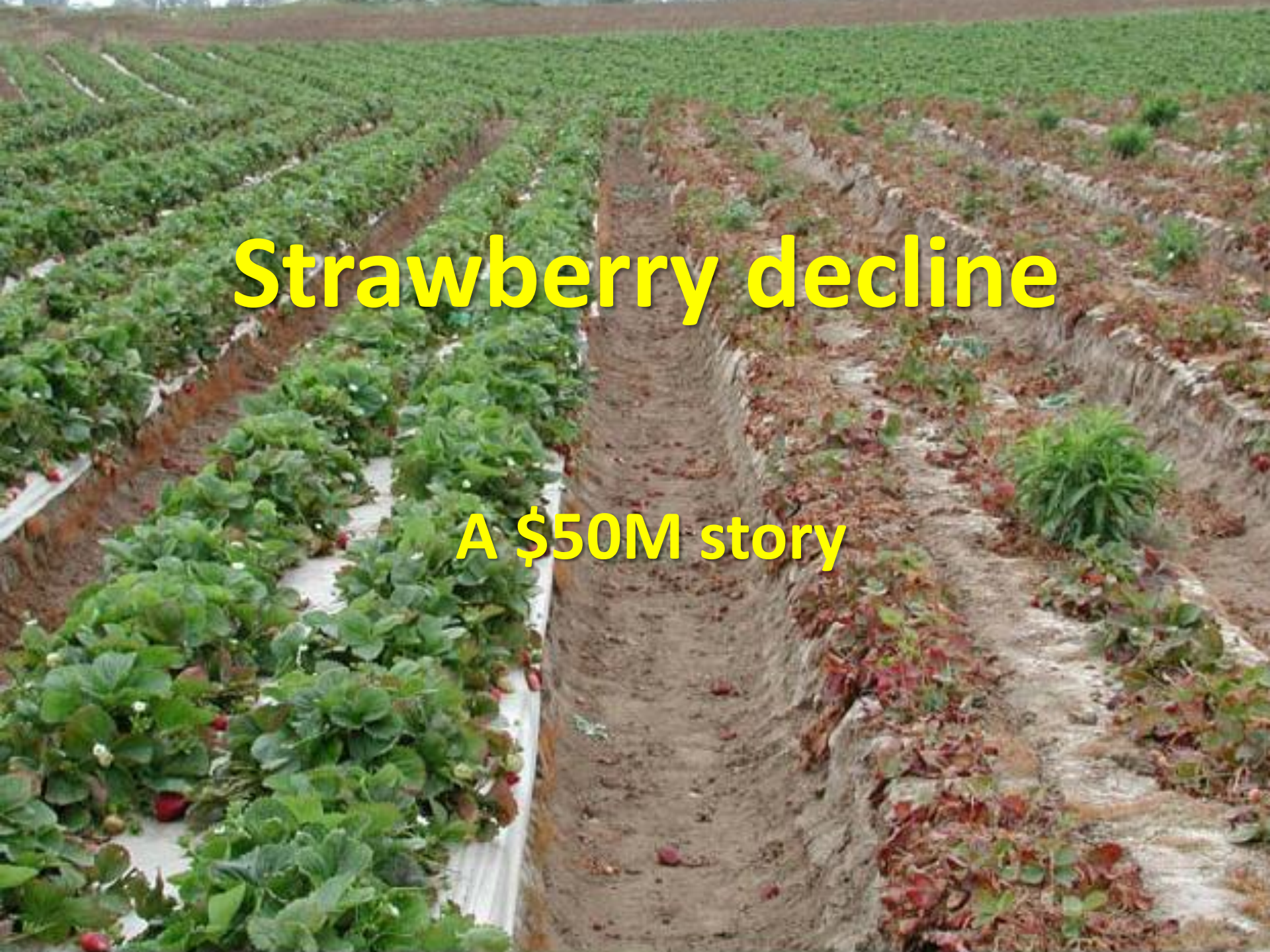
Strawberry pallidosis

Beet pseudo-yellows

Whiteflies*

Strawberry decline

A \$50M story







The whitefly viruses

Table 2. Transmission frequency of Strawberry pallidosis-associated virus (SPaV) and Beet pseudo yellows virus (BPYV) with the greenhouse whitefly (*Trialeurodes vaporariorum*)

Virus	Plant species	
	<i>Fragaria</i> × <i>ananassa</i>	<i>Nicotiana</i> <i>benthamiana</i>
BPYV	8/21 (38%)	16/20 (80%)
SPaV	3/21 (14%)	3/20 (15%)

Table 3. Host range of Strawberry pallidosis-associated virus (SPaV) utilizing *Trialeurodes vaporariorum* for transmission

Plant species tested	Infected/ inoculated
<i>Fragaria</i> × <i>ananassa</i>	3/10
<i>Sibbaldia procumbens</i>	3/8
<i>Duchesnea indica</i>	0/3
<i>Nicotiana benthamiana</i>	6/10
<i>N. glutinosa</i>	0/5
<i>N. clevelandii</i>	2/5
<i>N. tabacum</i>	0/5
<i>Physalis wrightii</i>	5/6
<i>P. floridana</i>	0/5
<i>Malva parviflora</i>	2/5
<i>Citrullus</i> spp.	0/5
<i>Chenopodium murale</i>	0/5
<i>C. capitatum</i>	0/5
<i>C. amaranticolor</i>	0/5
<i>Gomphrena globosa</i>	0/5
<i>Capsella bursa-pastoris</i>	0/5
<i>Brassica oleracea</i> var. <i>italica</i>	0/5
<i>Lycopersicon esculentum</i>	0/5
<i>Beta vulgaris</i>	0/5
<i>B. maritima</i> subsp. <i>macrocarpa</i>	0/5
<i>Datura stramonium</i>	0/5
<i>Urtica urens</i> ^a	0/5

^a SPaV has also been found in field isolates of nettle (*Urtica* sp.) associated with high field populations of greenhouse whitefly. It is not known if field isolates were *U. urens* or another *Urtica* species.

BPYV-SPaV double infections



Other strawberry viruses?

Revisited strawberry virus-like diseases.



Chlorotic fleck



Leafroll

Goal: Identify unknown viruses that may contribute to the decline.

Identification, characterization and development of detection techniques for strawberry viruses

Virus name	Acronym	Mode of transmission	Genus	Laboratory detection ^b
* <i>Apple mosaic</i>	ApMV	Pollen, seed	<i>Ilarvirus</i>	ELISA, RT-PCR
<i>Arabis mosaic</i>	ArMV	Nematode, seed	<i>Nepovirus</i>	ELISA, RT-PCR
* <i>Beet pseudo-yellow</i>	BPYV	Whitefly	<i>Crinivirus</i>	RT-PCR
* <i>Fragaria chiloensis cryptic</i>	FCICV	Unknown	Unknown	RT-PCR
* <i>Fragaria chiloensis latent</i>	FCILV	Pollen, seed	<i>Ilarvirus</i>	ELISA, RT-PCR
<i>Raspberry ringspot</i>	RpRSV	Nematode, seed	<i>Nepovirus</i>	ELISA, RT-PCR
* <i>Strawberry chlorotic fleck</i>	StCFV	Aphid	<i>Closterovirus</i>	RT-PCR
<i>Strawberry crinkle</i>	SCV	Aphid	<i>Cytorhabdovirus</i>	RT-PCR
Strawberry feather leaf	NA	Unknown	Unknown	NA
* <i>Strawberry latent</i>	StLV	Unknown	<i>Cripavirus</i>	RT-PCR
<i>Strawberry latent C</i>	SLCV	Aphid	<i>Nucleorhabdovirus</i>	NA
* <i>Strawberry latent ringspot</i>	SLRSV	Nematode, seed	<i>Sadwavirus</i>	ELISA, RT-PCR
<i>Strawberry mild yellow edge</i>	SMYEV	Aphid	<i>Potexvirus</i>	ELISA, RT-PCR
<i>Strawberry mottle</i>	SMoV	Aphid	<i>Sadwavirus</i>	RT-PCR
* <i>Strawberry necrotic shock</i>	SNSV	Thrips, pollen, seed	<i>Ilarvirus</i>	ELISA, RT-PCR
* <i>Strawberry pallidosis associated</i>	SPaV	Whitefly	<i>Crinivirus</i>	RT-PCR
<i>Strawberry pseudo mild yellow edge</i>	SPMYEV	Aphid	<i>Carlavirus</i>	ELISA
<i>Strawberry vein banding</i>	SVBV	Aphid	<i>Caulimovirus</i>	PCR
<i>Tobacco necrosis</i>	TNV	Oomycete	<i>Necrovirus</i>	ELISA, RT-PCR
<i>Tomato black ring</i>	TBRV	Nematode, seed	<i>Nepovirus</i>	ELISA, RT-PCR
<i>Tomato ringspot</i>	ToRSV	Nematode, seed	<i>Nepovirus</i>	ELISA, RT-PCR

Tobacco streak virus

* *Strawberry crinivirus 3*

* *Strawberry crinivirus 4*

No vector control



Vector control



Blackberry yellow vein disease

First observed in 2000 in the Carolinas.



Tested for known viruses (RBDV, TRSV etc) – Several viruses were found but none consistently associated with symptoms.

Tobacco ringspot virus and BYVD

BYVD is very similar to what people thought as being TRSV symptoms

TRSV textbook symptoms



Single TRSV-infection



Are symptoms cv. dependent? The **majority** of plants infected with TRSV are symptomless

New viruses in *Rubus* in the last 7 yrs

16 viruses & virus-like agents were known to infect *Rubus* before we started looking into *Rubus* complexes – We now have over 40...

New *Rubus* viruses

Blackberry yellow vein associated virus

Blackberry virus E

Blackberry virus X

Blackberry virus Y

Blackberry virus Z

Beet pseudo yellows virus

Blackberry yellow mottle virus

Blackberry chlorotic ringspot virus

Strawberry necrotic shock virus

Black raspberry necrosis virus

Raspberry leaf mottle virus

Rubus canadensis virus -1

Impatiens necrotic spot virus

Raspberry latent virus

etc.....

New viruses in *Rubus* in the last 8 yrs

16 viruses & virus-like agents were known to infect *Rubus* before we started looking into *Rubus* complexes – We now have over 40...

New *Rubus* viruses

Blackberry yellow vein associated virus

Blackberry virus E

Blackberry virus X

Blackberry virus Y

Blackberry virus Z

Beet pseudo yellows virus

Blackberry yellow mottle virus

Blackberry chlorotic ringspot virus

Strawberry necrotic shock virus

Black raspberry necrosis virus

Raspberry leaf mottle virus

Rubus canadensis virus -1

Impatiens necrotic spot virus

Raspberry latent virus

etc.....

Tests are available for all the new viruses

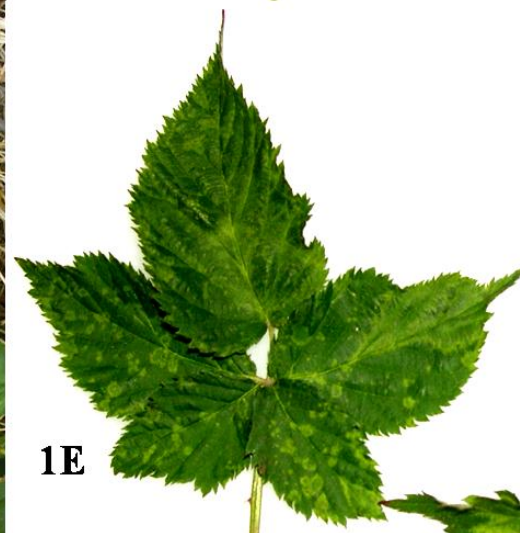


Same disease-different viruses

AR



MS



NC



Arkansas

BYVaV

BVY

TRSV



Carolinas

BYVaV

BVX

BPYV

INSV

TRSV



Mississippi

BYVaV

TRSV

BVE



How do we tackle BYVD?

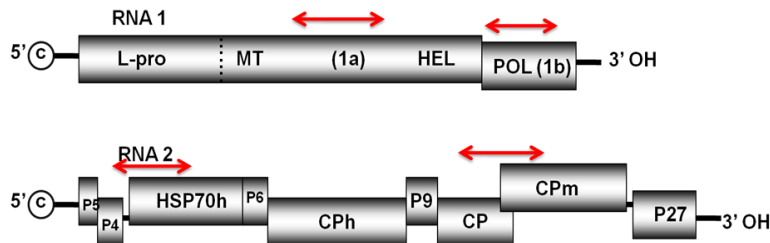
After identification of all (or almost all) viruses that are involved in the disease we need to:

- A. Make sure that mother plants are being tested for the new viruses before they are propagated.**
- B. Identify virus combinations that can cause BYVD.
- C. Identify virus vectors.
- D. Find alternative hosts of the viruses in the field.
- E. Minimize or eliminate BYVD by eliminating the weakest link, the virus vector(s) that is the easiest to control.

What are the viruses present in your area?

The importance of detection

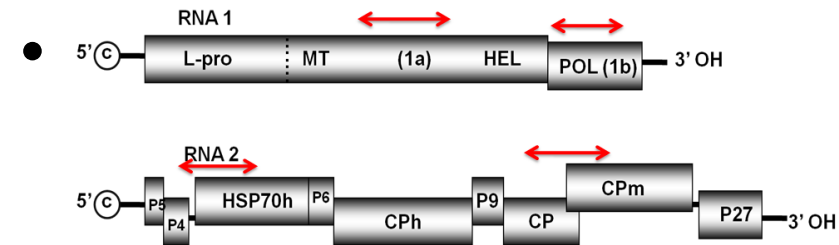
- BYVaV - Multistate sample collection - 35 isolates



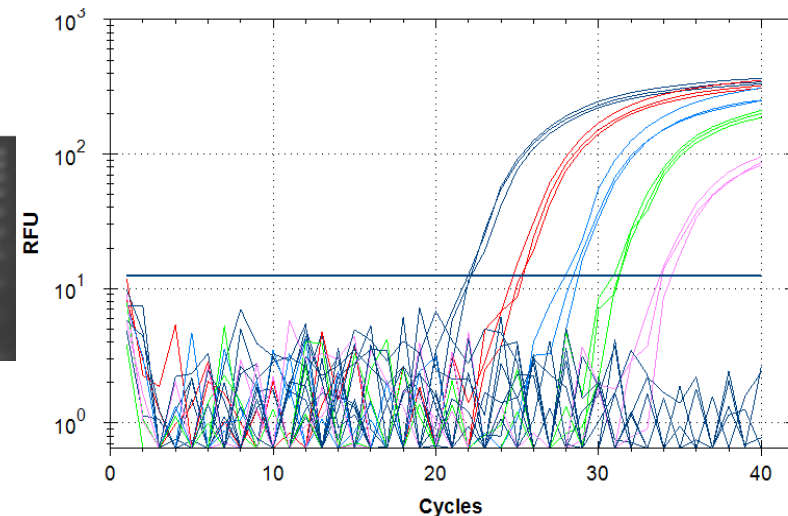
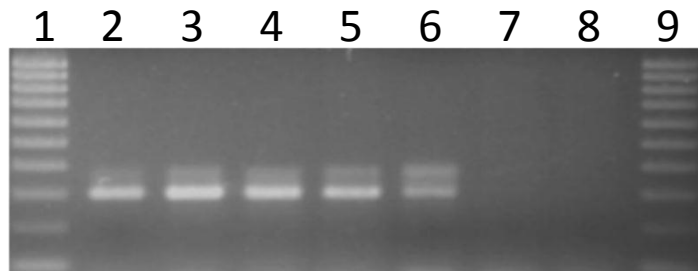
What are the viruses present in your area?

The importance of detection

- BYVaV - Multistate sample collection - 35 isolates



- Detection
100%
identity



Virus interactions: The BYVaV/BVY story

BVY did not cause symptoms in single infections but together with BYVaV they cause BYVD.

In mixed infections, BVY knocks down concentration of BYVaV to about 0.1% compared to titer in single infections.

In mixed infections, they can cause death of fruiting canes.

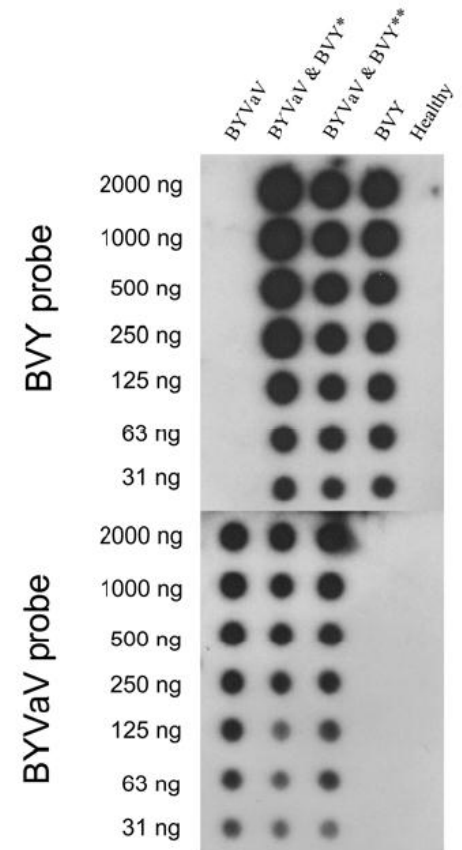


Fig. 4. Nucleic acid spot hybridization (NASH) detection of Blackberry virus Y (BVY) and Blackberry yellow vein associated virus (BYVaV) in blackberry. The blot was initially hybridized with a BVY-specific DNA probe and subsequently with a BYVaV-specific probe after stripping the BVY probe. *symptomatic mixed infection and **asymptomatic mixed infection.

Transmission

Experiment	<i>Trialeurodes abutilonea</i>	<i>Trialeurodes vaporariorum</i>
Experiment 1	4/7	3/9
Experiment 2	5/8	1/8
Experiment 3	3/10	3/10
Total	12/25	7/27



www.apsnet.org

Greenhouse whitefly

- Both whitefly species transmitted the virus at a rate >30%

Alternate hosts

Plant species	Scientific name	Family	Number of plants tested
Garden vetch	<i>Vicia sativa</i>	Fabaceae	16
Virginia creeper	<i>Parthenocissus quinquefolia</i>	Vitaceae	16
Red clover	<i>Trifolium pretense</i>	Fabaceae	16
Wild garlic	<i>Allium vineale</i>	Amaryllidaceae	16
Creeping woodsorrel	<i>Oxalis corniculata</i>	Oxalidaceae	16
Carolina geranium	<i>Geranium carolinianum</i>	Geraniaceae	16
Curly dock	<i>Rumex crispus</i>	Polygonaceae	16
Dandelion	<i>Taraxacum officinale</i>	Asteraceae	16
Tall fescue	<i>Festuca arundinacea</i>	Poaceae	16
Wild wheat	<i>Avena fatua</i>	Poaceae	16
Grapes	<i>Vitis vinifera</i>	Vitaceae	16
Peach	<i>Prunus persica</i>	Rosaceae	16
Blueberry	<i>Vaccinium</i> spp.	Ericaceae	16
Shepherd's purse	<i>Capsella bursa-pastoris</i>	Brassicaceae	16
Nutsedge	<i>Cyperus</i> spp.	Cyperaceae	16
Horsenettle	<i>Solanum carolinense</i>	Solanaceae	16
Common ragweed	<i>Ambrosia artemisiifolia</i>	Asteraceae	16
Tree of heaven	<i>Ailanthus altissima</i>	Simaroubaceae	16
Apple	<i>Malus</i> spp.	Rosaceae	200
Rose	<i>Rosa multiflora</i>	Rosaceae	40
Carpetweed	<i>Mollugo verticillata</i>	Molluginaceae	16
Amaranthus	<i>Amaranthus</i> spp.	Amaranthaceae	16
Poor joe	<i>Diodia teres</i>	Rubiaceae	16
Ground cherry	<i>Physalis</i> spp.	Solanaceae	16
Sorghum	<i>Sorghum</i> spp.	Poaceae	16



Vector elimination

The BRNV paradigm

New field monitoring

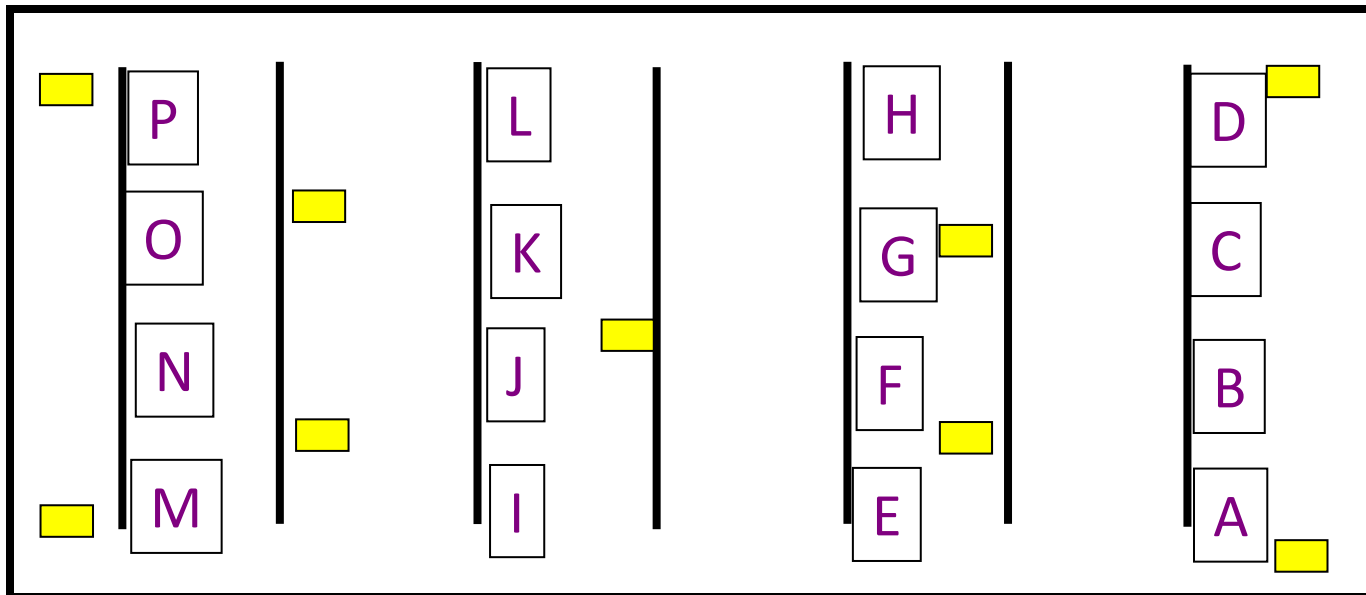
Permanent tagged plants

Time of transmission

High incidence of virus

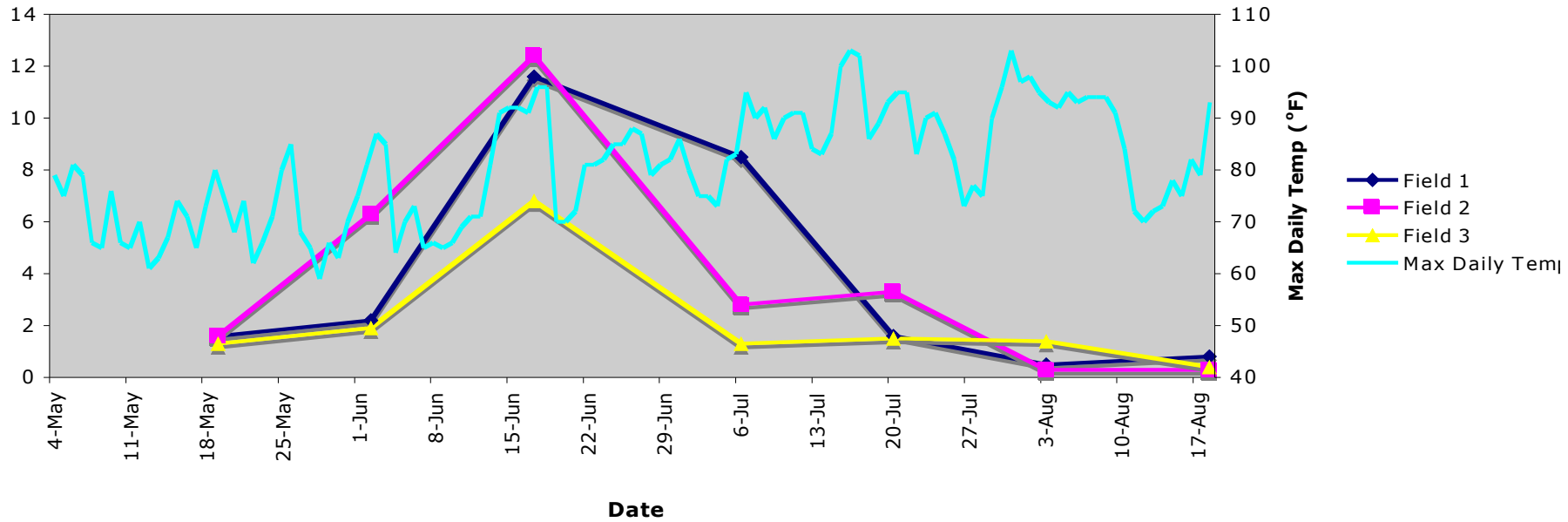
Potted plants

Rotated every month

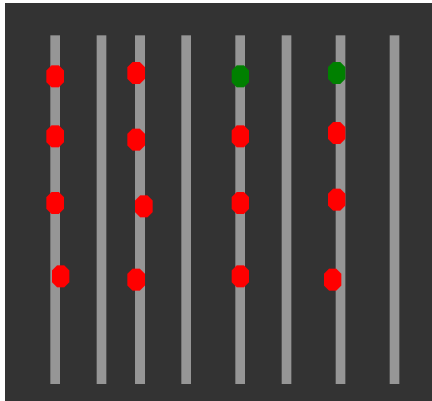




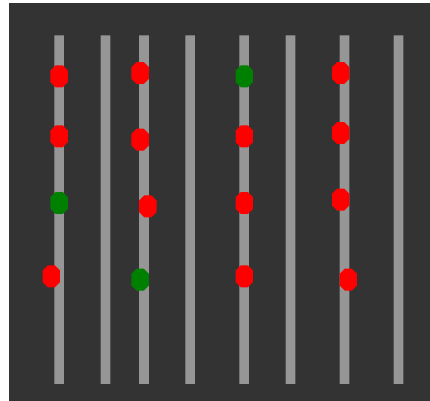
Average Aphids/Trap:



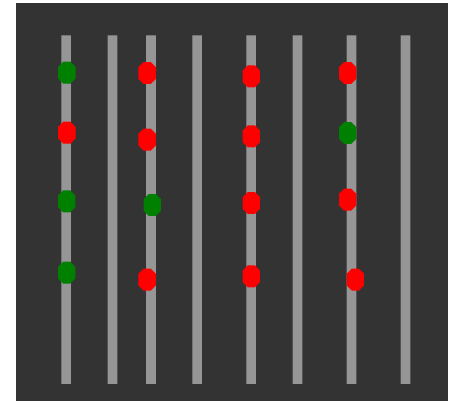
Field 1

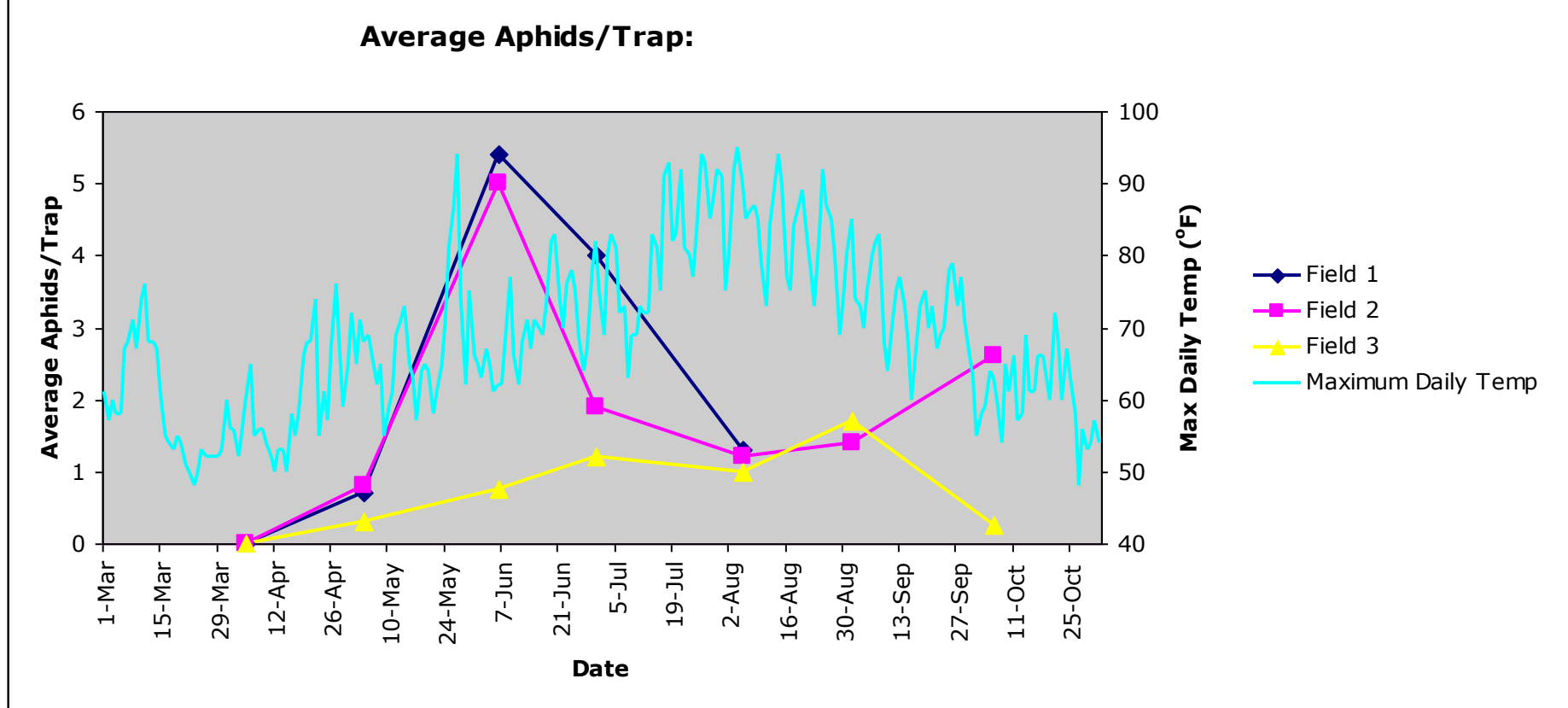


Field 2

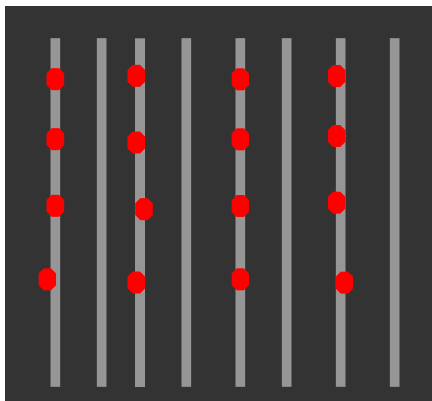


Field 3

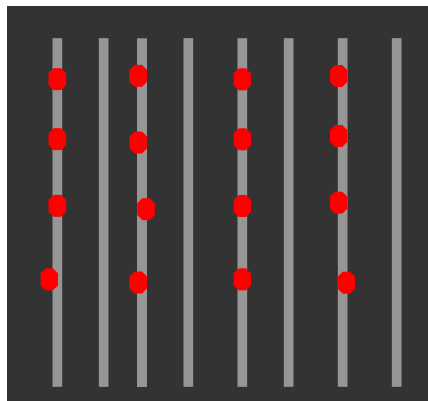




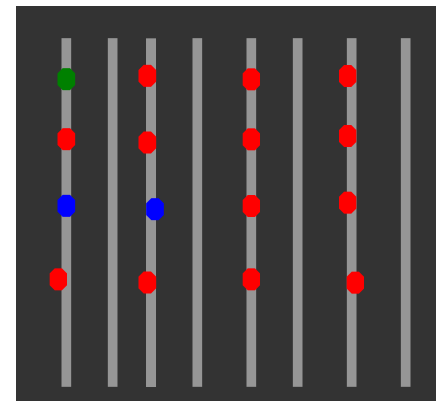
Field 1



Field 2



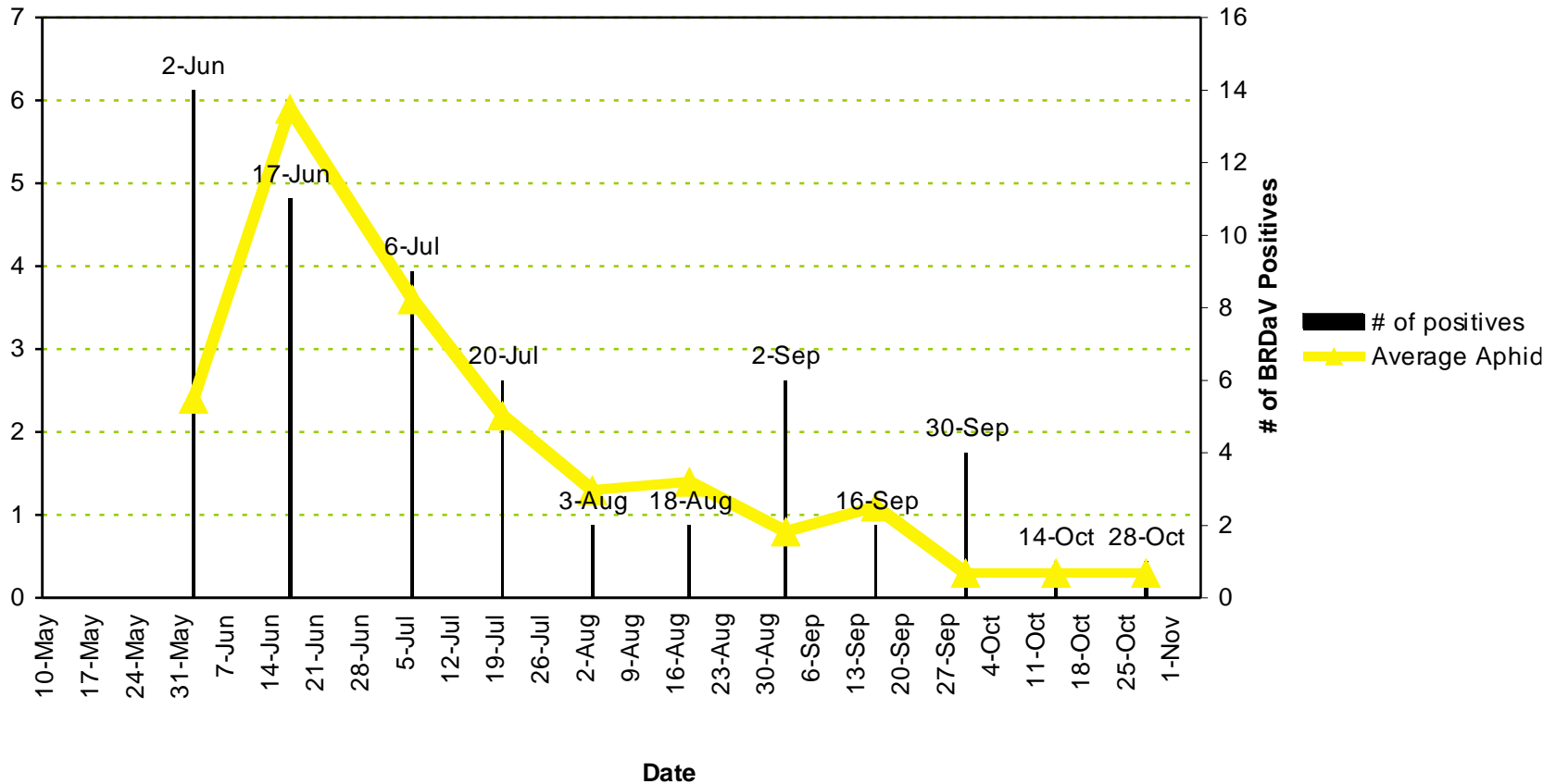
Field 3



Nearly 100% transmission in three years!

Time of Transmission

Average Aphids/Trap vs. Positives:

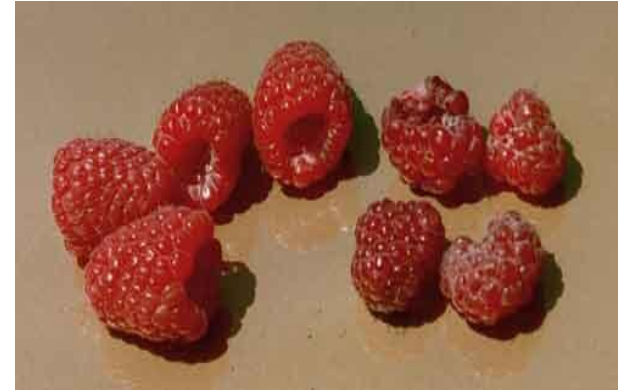


Raspberry crumbly fruit and decline

- The Pacific Northwest (PNW) is a primary producer of red raspberries

Raspberry crumbly fruit and decline

- 'Several cultivars are susceptible to crumbly fruit disease (drupelets abortion)
- *Raspberry bushy dwarf virus* (RBDV), a pollen-borne ideovirus was considered the causal agent of crumbly fruit
- Still, in many cases RBDV single infections did not cause symptoms



Another virus complex?

Important observations suggested that crumbly fruit symptoms may be increased by additional viruses:

1. The disorder is more severe in cool areas with high populations of the large raspberry aphid *Amphorophora agathonica*



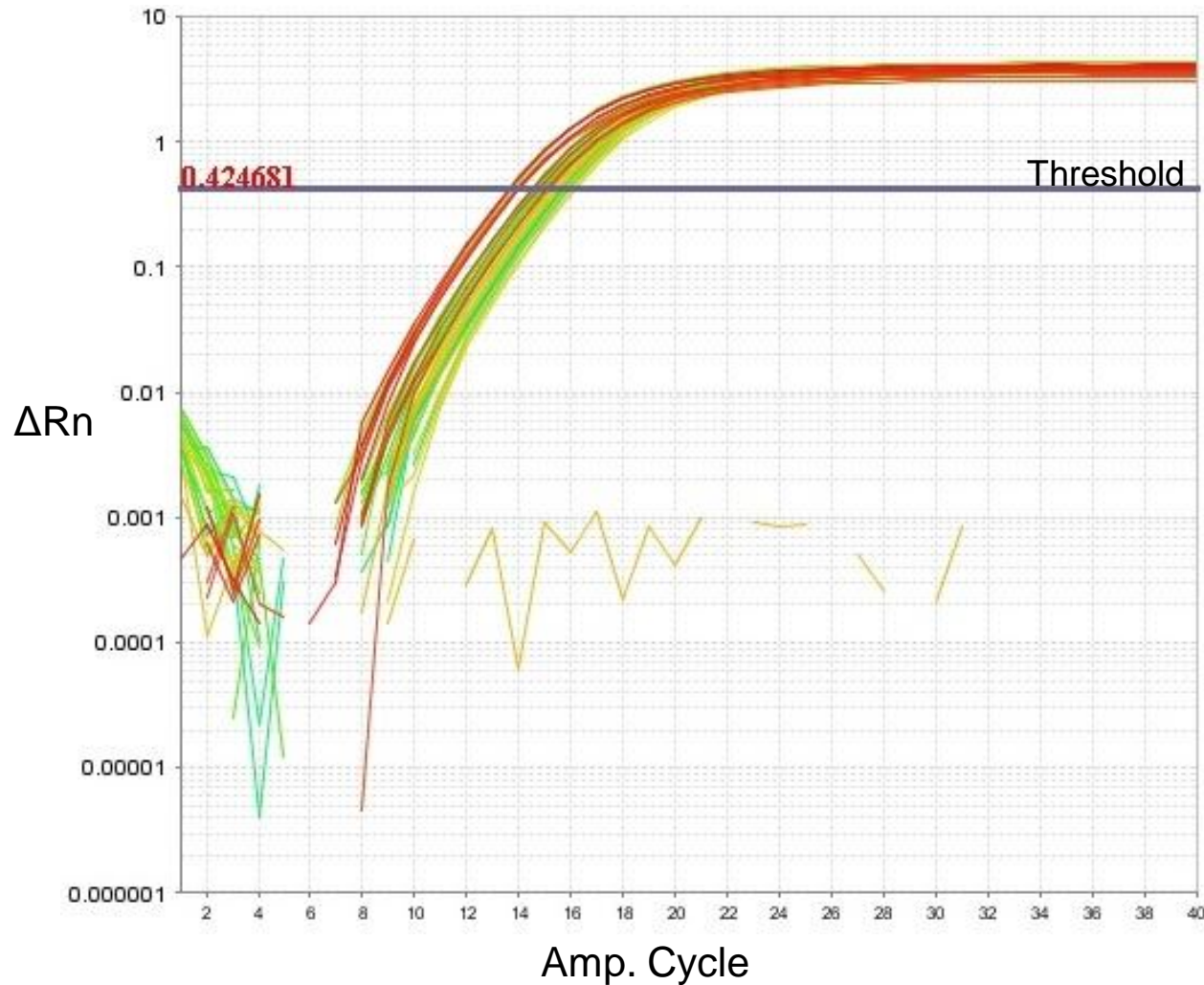
2. Two additional viruses found in severely affected fields, **Raspberry leaf mottle (RLMV)** and **Raspberry latent (RpLV)**

RBDV, RLMV and RpLV interactions



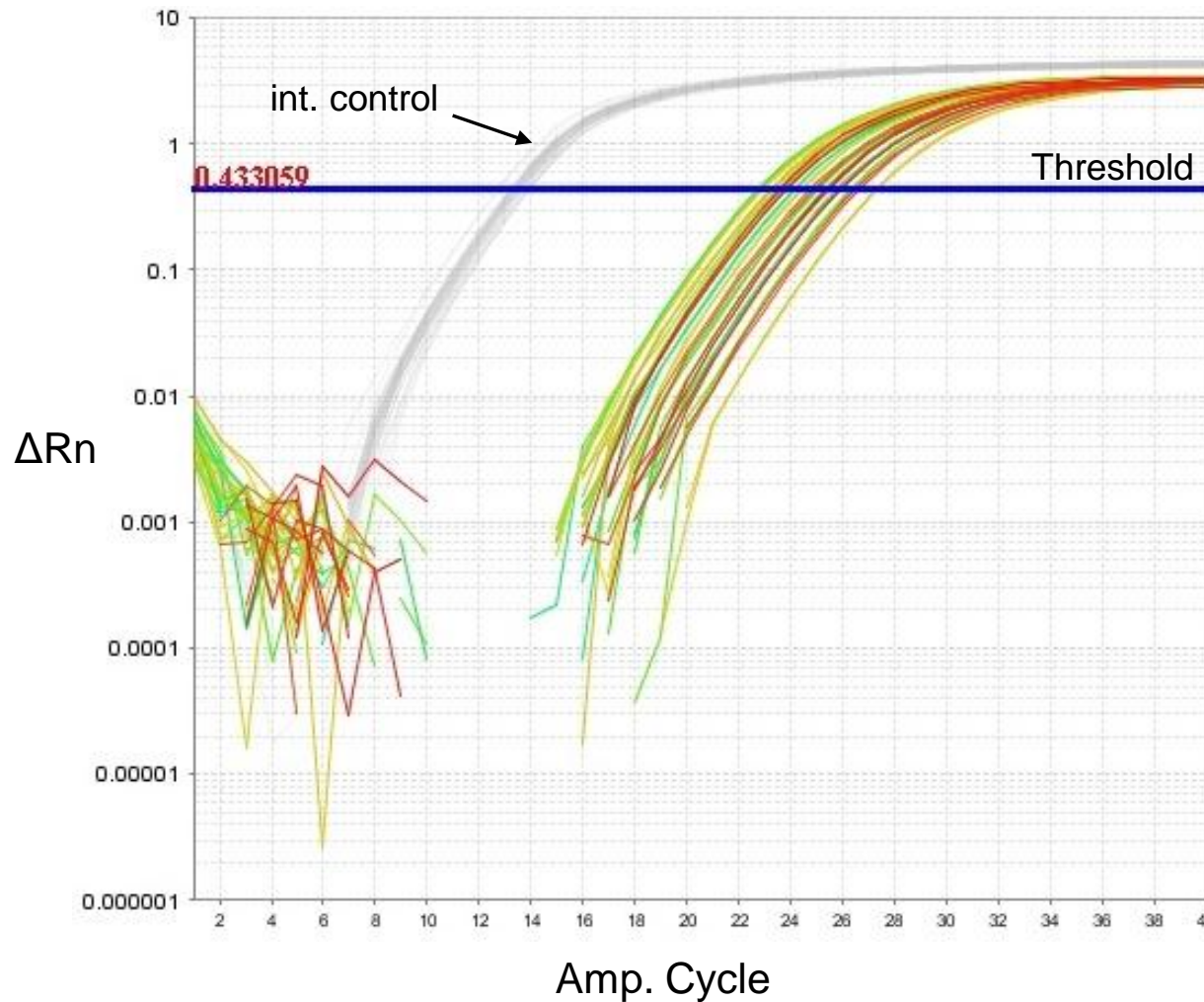
RLMV qRT-PCR

RLMV titer in single and mixed infections over time

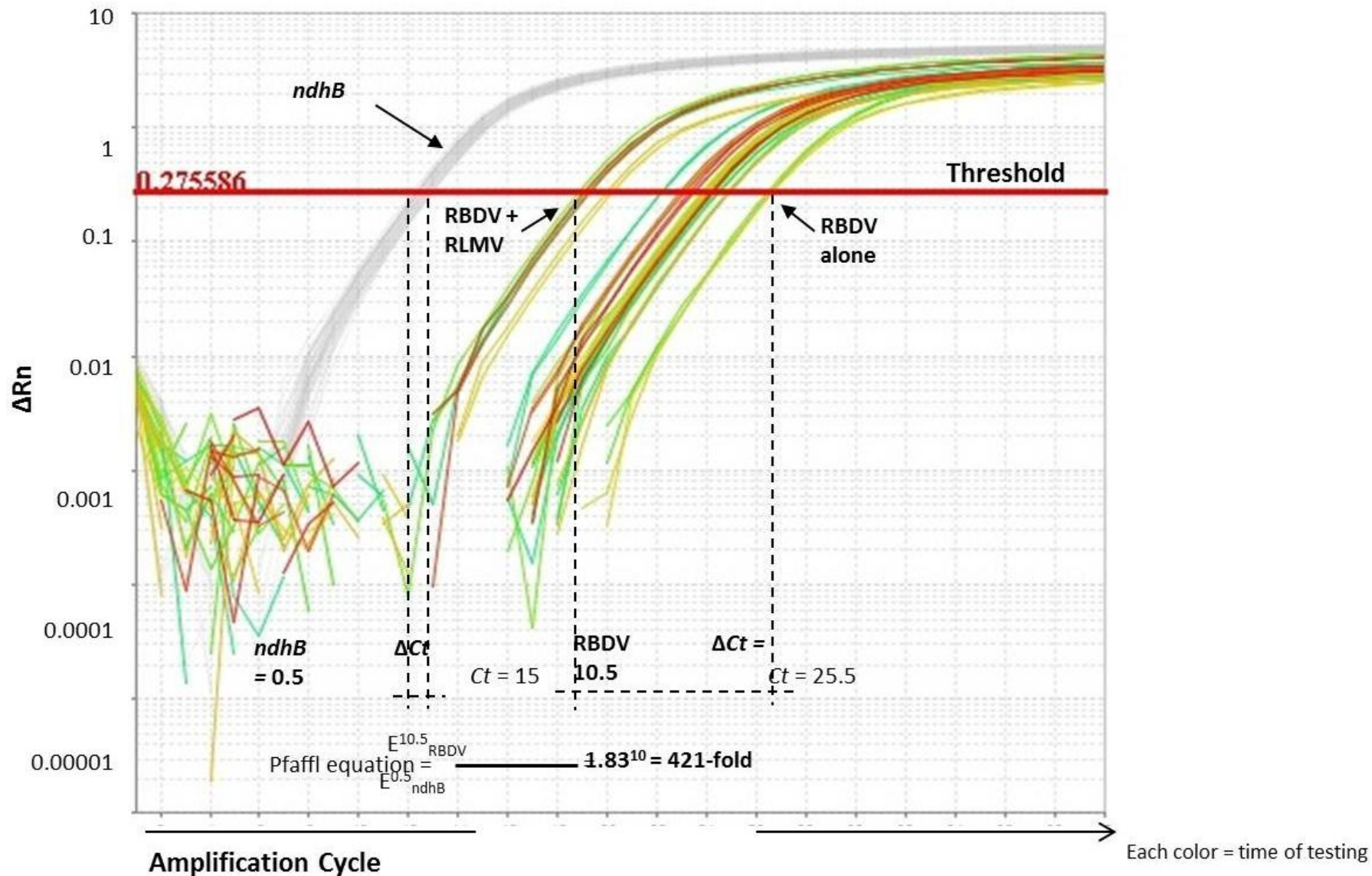


RpLV qRT-PCR

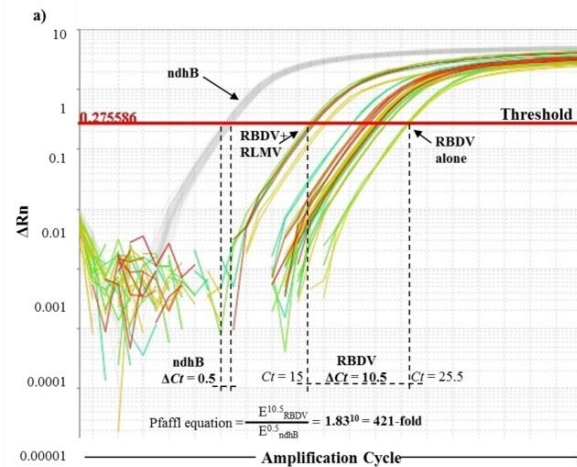
RpLV titer in single and mixed infections over time



RBDV titer enhanced in co-infections with RLMV

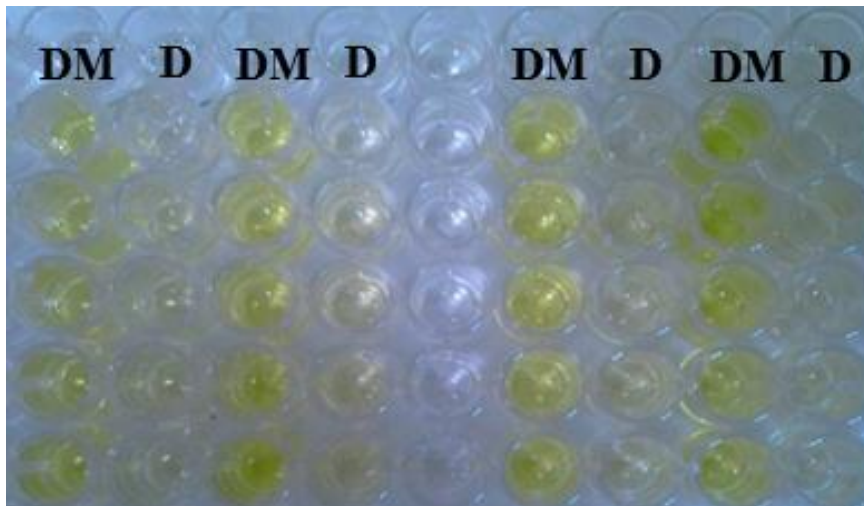


RBDV titer enhanced in co-infections with RLMV

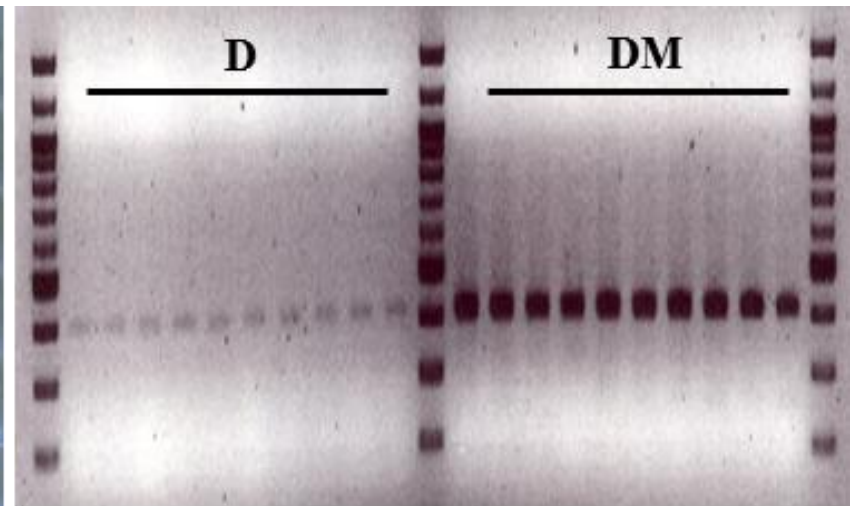


RBDV titer increase verified by conventional methods

ELISA

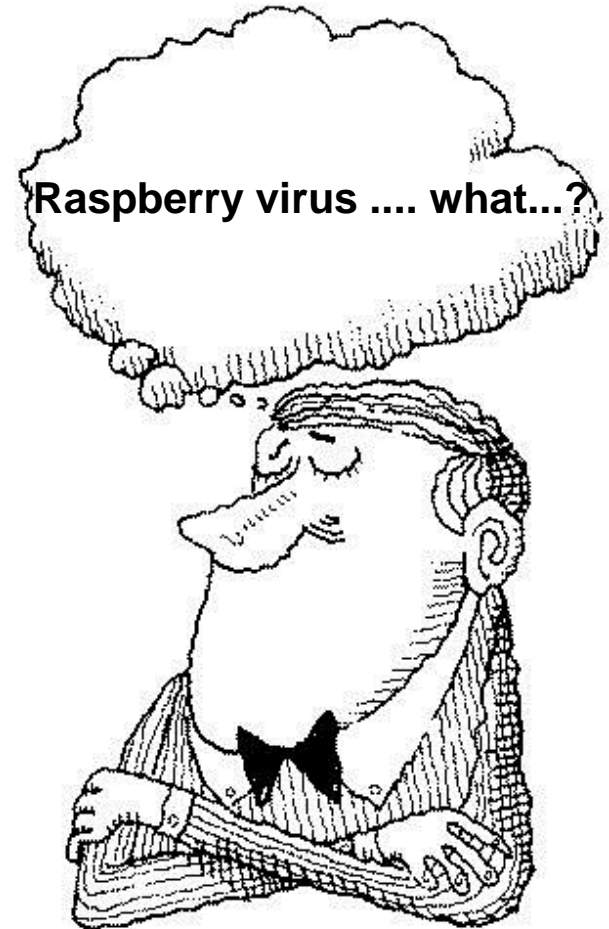


RT-PCR (20 cycles)



Mixed virus infections affect on plant growth and fruit crumbliness

- **H** **Control**
- **D** **RBDV - Dwarf**
- **M** **RLMV - Mottle**
- **L** **RpLV - Latent**
- **DM** **RBDV + RLMV**
- **DL** **RBDV + RpLV**
- **ML** **RLMV + RpLV**
- **DML** **RBDV + RLMV + RpLV**

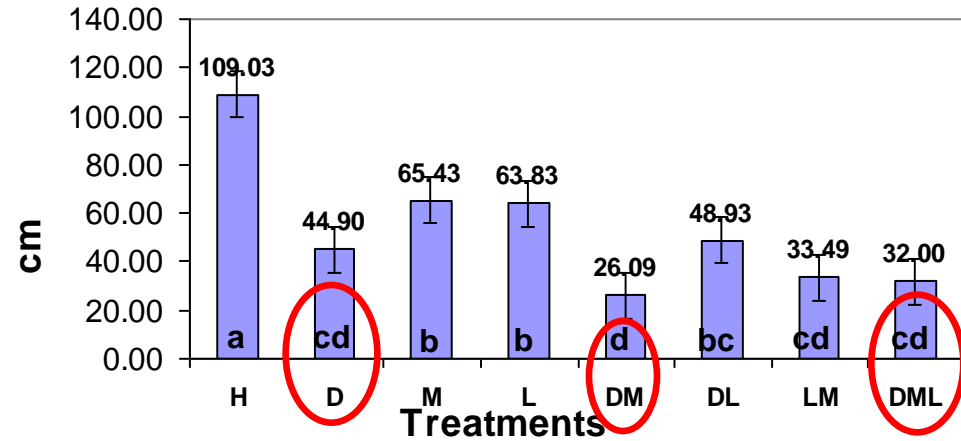


Plant Growth Establishment (2010)

Oct.



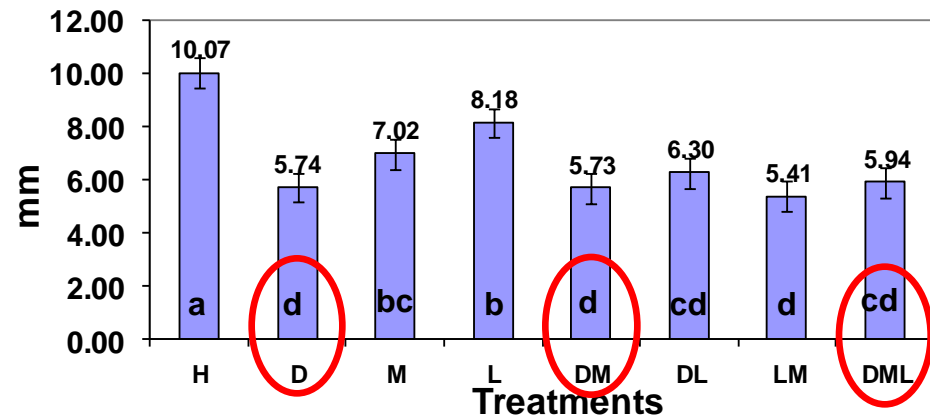
Height



Oct.



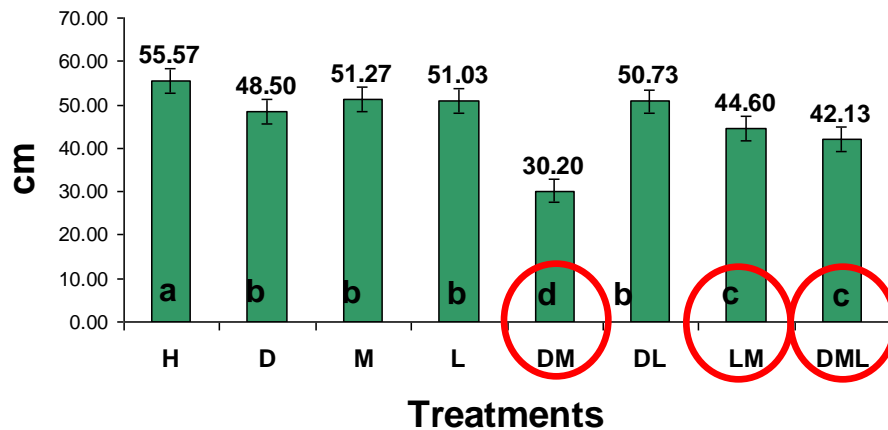
Cane diameter



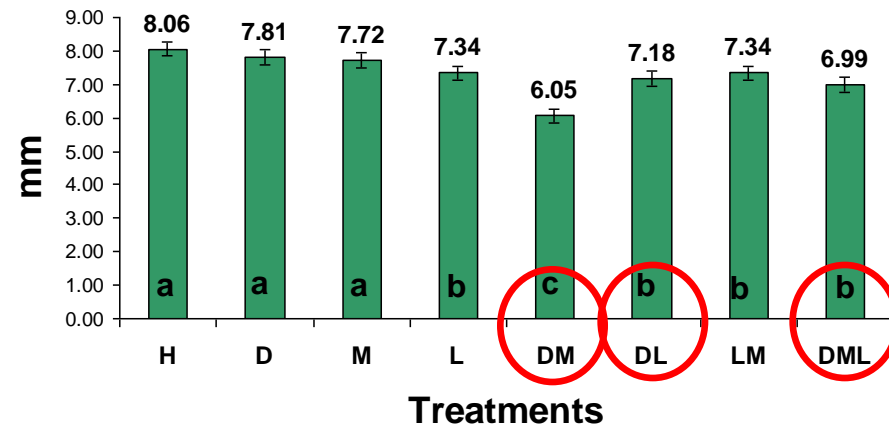
Plant Growth (2011)



Height



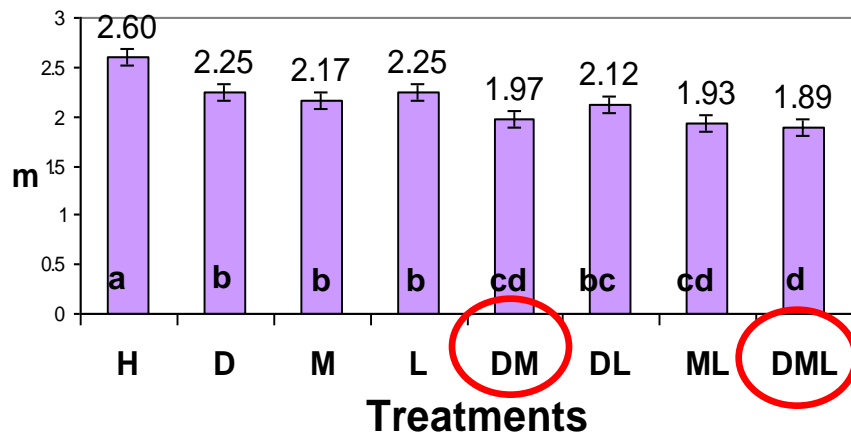
Cane diameter



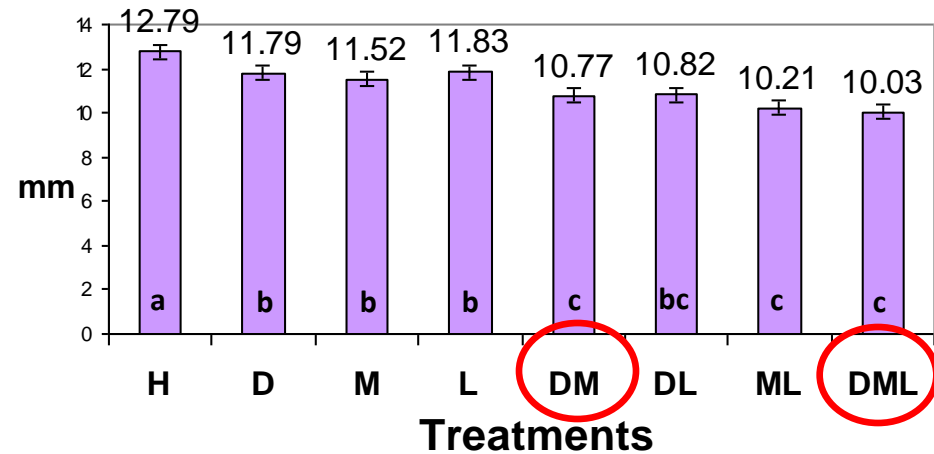
Plant Growth (2011)



Height

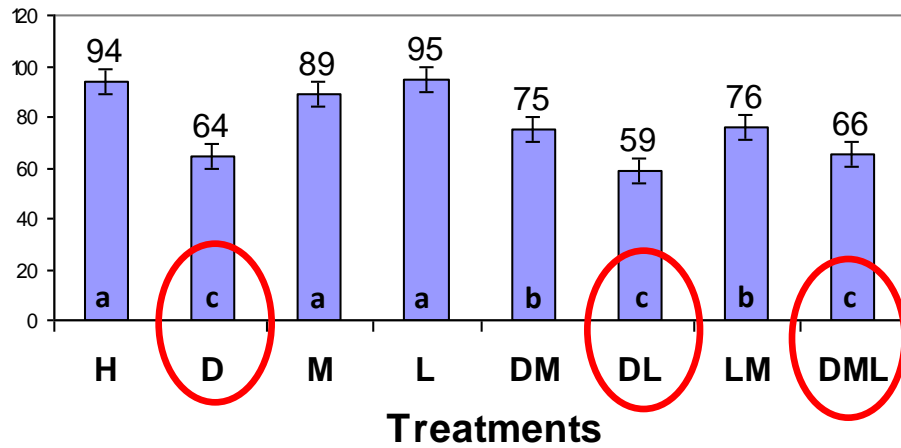


Cane diameter

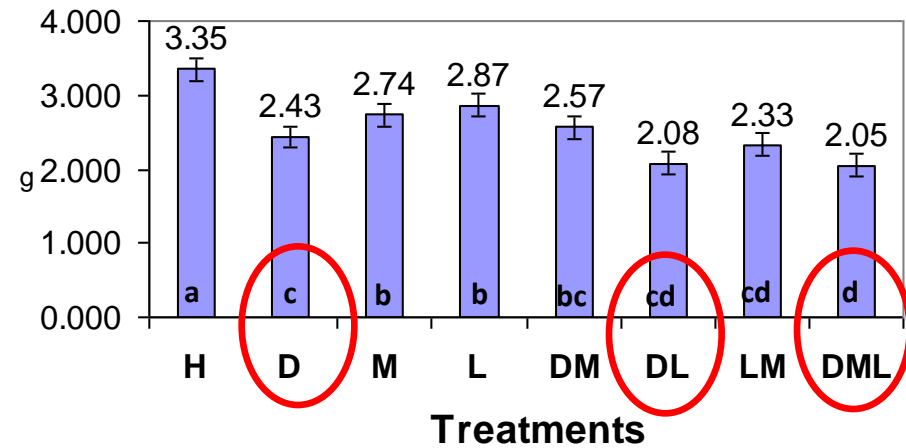


Crumbly Fruit

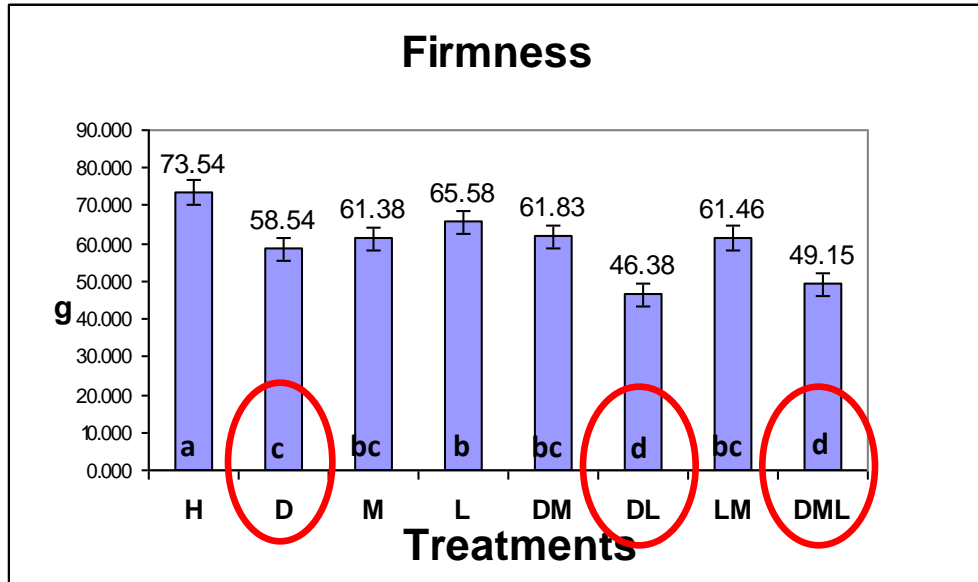
No. Drupelets



Weight



Crumbly fruit



Virus Incidence in 'Meeker' Fields

Northern Washington		
Field Age (years)	RLMV (%)	RpLV (%)
1	4	0
1	30	0
1	10	0
2	58	21
2	0	0
2	0	0
2	6	0
2	16	0
3	31	6
3	6	0
3	13	0
3	50	0
4	19	6
4	13	0
5	69	0
5	90	80
5	100	75
5	44	6
5	100	17
6	70	25
6	100	6
6	100	12
7	100	6
8	100	46

Southern Washington/Oregon		
Field Age (years)	RLMV (%)	RpLV (%)
1	0	0
5	40	20
6	0	20
7	8	17
8	19	0
8	27	0



Crumbly Fruit Scouting

Crumbly fruit and virus incidence in Washington

Field Age	Crumbliness 0: normal 3: severe	Virus incidence %		
		RBDV	RLMV	RpLV
	0 1 2 3			
4	1	44	25	0
4	2	100	100	0
4	3	93	100	7
5	3	100	100	40
6	3	92	96	40



Insects in Traps (2011)

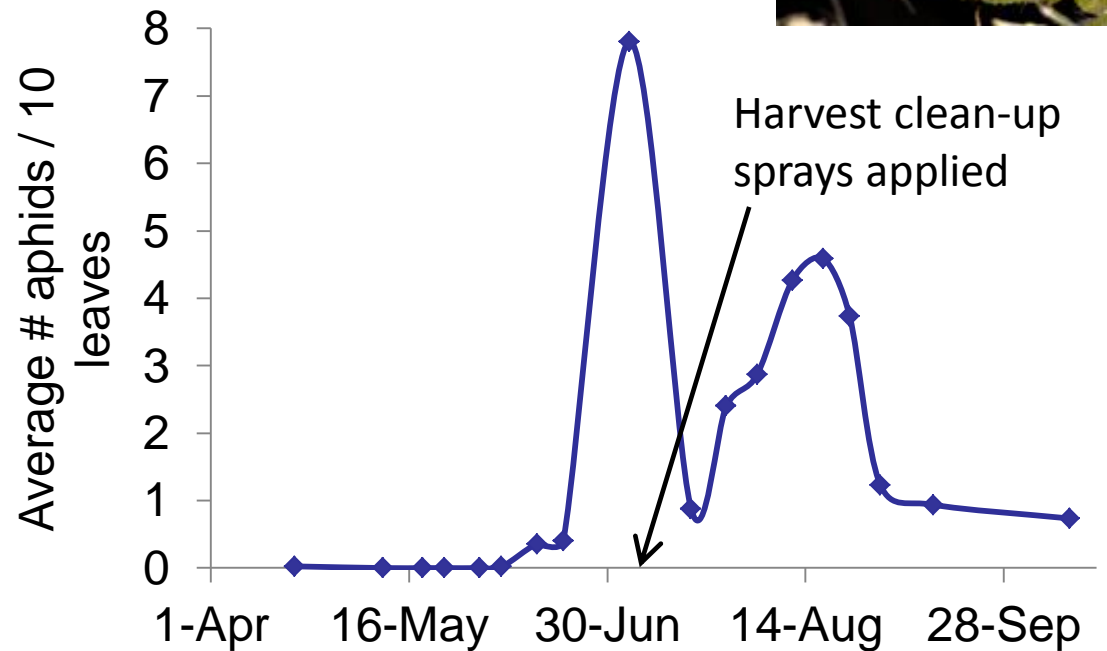
Empoasca fabae was sporadic



Few numbers of *Macropsis fuscula*



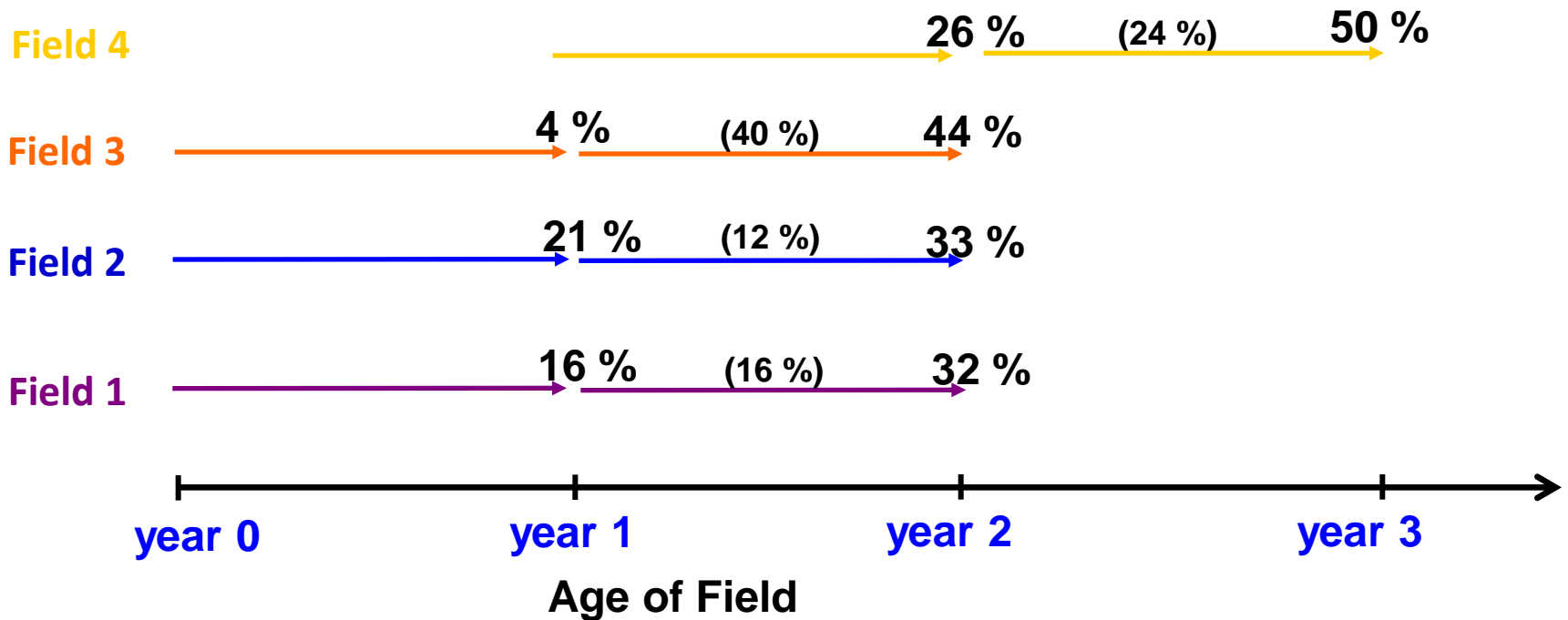
Raspberry aphid *A. agathonica*
predominant insect



Lightle, unpublished data

RLMV spread in the field

Four fields being monitored for virus spread



Control Strategies

1. Think long term, identify potential risks of a site
- 2. Start with clean plants**
3. Identify and diagnose problems early
4. Implement control strategies ASAP
5. If a virus complex is involved - identify viruses present and which are the easiest to control

The importance of clean plants

- Better establishment



The importance of clean plants

- Better establishment
- Longer life of plantings



The importance of clean plants

- Better establishment
- Longer life of plantings
- Fewer disease problems/Reduce risk of introducing new viruses to a region or field





National Clean Plant Network

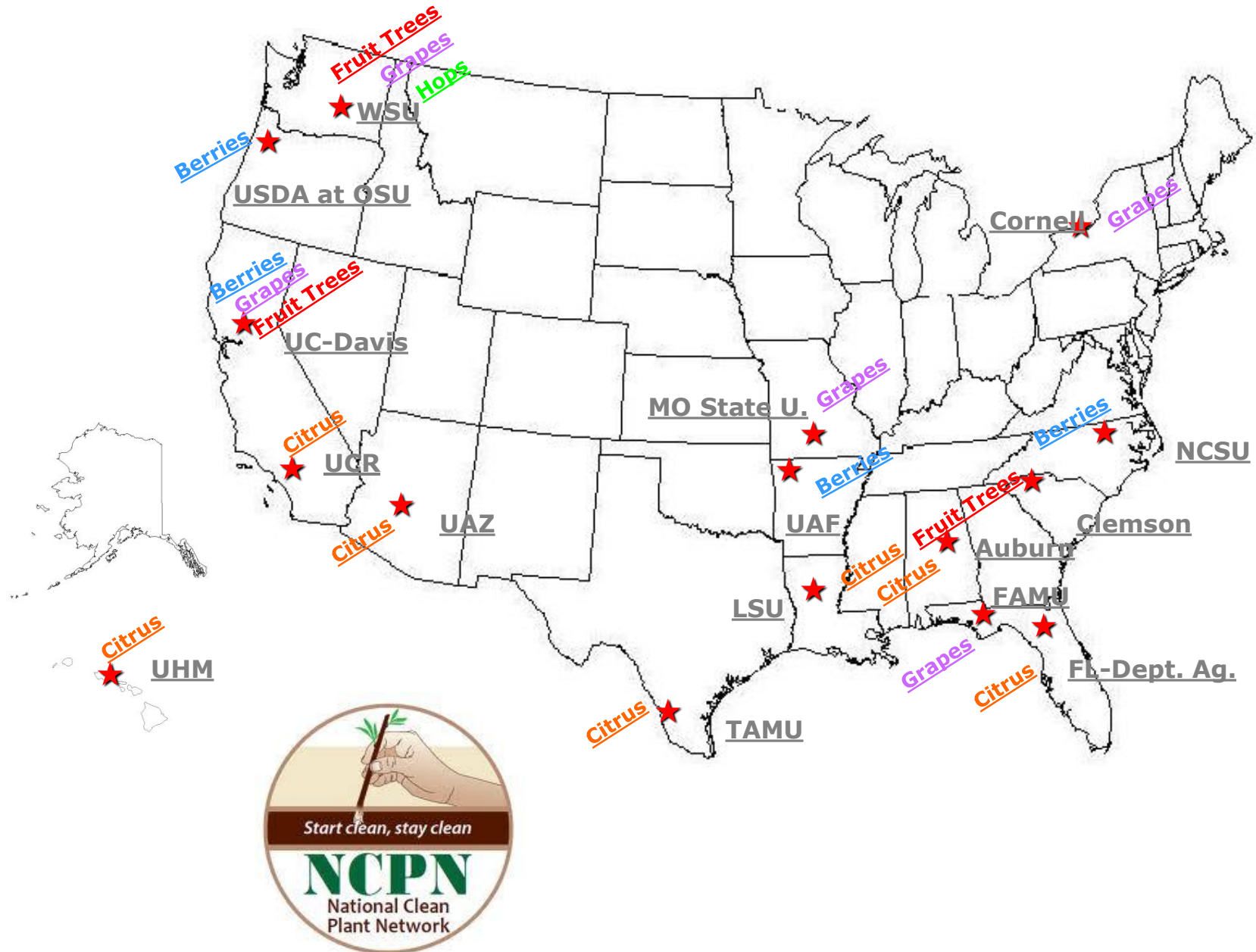


A federally-coordinated effort to secure high quality virus-tested plants for clonally propagated crops.

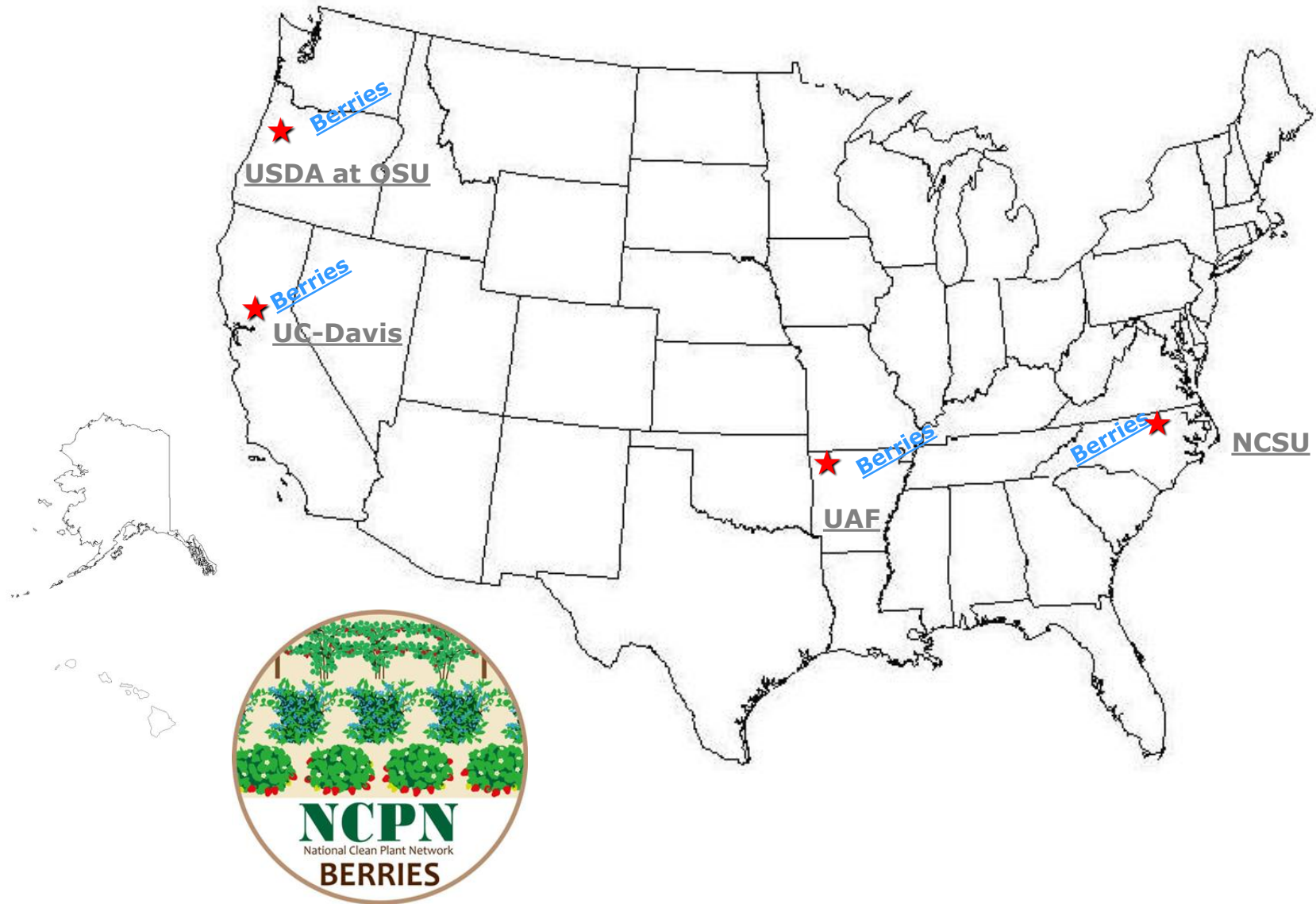
NCPN Mission

The NCPN provides high quality asexually propagated plant material free of targeted plant pathogens and pests that cause economic loss to protect the environment and ensure the global competitiveness of specialty crop producers in the United States.

NCPN Supported Clean Plant Centers



Berry Clean Plant Centers



Why care???



The story:

Propagation from an existing plot

10 ton/acre = \$30,000/year

Latent infections with Blueberry scorch

The result?

Removal of infected material

Cumulative loss: ~ 100,000/acre



Florida 2013







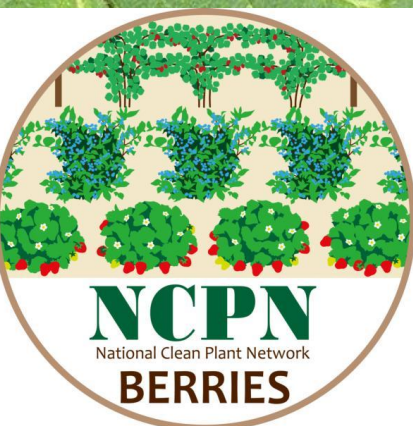
Start clean!!!!

The team

The berry virus consortium:

16 individuals from UA, NCSU, USDA-ARS, MSU, UGA

Bindu Poudel, Diego Quito, Danielle Lightle, Anne Halgren,
James Susaimuthu



United States Department of Agriculture
National Institute of Food and Agriculture

